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PRESSURE STUDIES OF PROTEIN DYNAMICS

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Annual Report to Office of Naval Research



1. Summary of Research Goals and Methods

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The key goal of biophysical and biochemical research is a fundamental understanding of the relation among protein structure, motions, and function (1). We have developed a hierarchical model to describe protein conformations and motions (2,3). We study the binding of small ligands like carbon monoxide (CO) and dioxygen to myoglobin (Mb) and other heme proteins. Our main experimental tool is flash photolysis over wide ranges of time, temperature, and pressure as well as solvent viscosity and pH (4,5). Studies of ligand binding to myoglobin over wide ranges of pressure and temperature probe protein states and motions which are not accessible by varying only temperature or pressure. Although living systems function over wide ranges in pressure from high altitude to the ocean depths, pressure effects have been relatively unexplored in protein science because of their complexity and subtlety (6,7,8). We have

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MbCO, for exploring the effects of pressure on proteins and protein reactions (9,10,11,12).

The concepts of conformational substates (13,14) and a slaved glass transition (10) in proteins are proving to be crucial in understanding the relation among protein structure, motions, and function. The concept of conformational substates implies that the ground state of a protein is highly degenerate and consists of many nearly isoenergetic minima in the conformational energy surface. A similar situation exists for spin glasses and other complex systems (15). The slaved glass transition (SGT) in myoglobin is similar to the "freezing" transition in a spin glass or glass (16,17). Our combined pressure and temperature experiments probe the glassy properties of proteins and, thus, connect two distant fields of research. This situation may lead to a more detailed understanding of both biomolecular relaxation and reactions and the physics of amorphous solids.

2. Summary of Accomplishments in Year Two

Significant progress has occurred during the second contract year. We have presented several papers on our results at meetings of the Biophysical Society (9,11,12). The work reported here has also resulted in one doctoral dissertation (18). We are preparing these results for a detailed publication. We only outline the main results in this report.

CO can bind to Mb in at least three positions which can be distinguished by their infrared CO stretching bands (19). In MbCO



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the three main stretching bands, denoted by A₀(~1966 cm⁻¹), A₁(~1945 cm⁻¹), and A₃(~1930 cm⁻¹), are sensitive to external influences such as pH, solvent. composition, temperature, and, most importantly for this project, pressure (9,10,11,12,18). Observation of the infrared band is done with two systems: (i) quasi-static and slow kinetics experiments are performed on a Fourier transform infrared (FTIR) spectrometer (Mattson Sirius 100 FTIR); (ii) fast kinetics experiments are performed on a homebuilt, microsecond, midinfrared flash photolysis system using a tunable infrared-diode to produce the monitoring light. We have also measured the heat capacity of the protein samples in a differential scanning calorimeter (11).

Two related classes of experiments were performed on MbCO: quasi-static and relaxation. In the quasi-static experiments we measure the characteristics of the CO stretching bands as function of temperature, pressure, pH, and solvent composition. We call the situation quasi-static because the glass-like behavior of the protein below a characteristic glass temperature Tg makes attainment of true equilibrium impossible on any reasonable timescale. In the relaxation experiments we observe the relaxation of the protein by monitoring the CO stretching bands after a fast pressure release at a temperature near to the glass temperature.

(a) Quasi-static Experiments. We have measured the IR spectrum of MbCO from 1910 to 1990 cm⁻¹ in a 75% glycerol/water solvent (pH 7 with potassium phosphate buffer) from 140K to 340K in temperature and 0.1 MPa to 100 MPa in pressure. (1 MPa = 10

bar). The spectra are resolved into three bands denoted in order of decreasing wavenumber by A₀, A₁, and A₃ which correspond to conformational substates of tier 0 also denoted by A₀ to A₃. The main results of the quasi-static experiments can be seen in Fig. 1 where we plot the temperature dependence of the ratio $r_0 = A_0/A_1$ of the total areas of the two bands at two different pressures. The ratio r₀ shows six different regions denoted by i-vi starting at low temperature: (i) Glass region. The protein is in a glass-like, nonequilibrium state in which large scale motions are frozen, but motions of groups of atoms still occur. This situation is dramatically illustrated by the temperature independence of the ratio r₀ over a timescale of several hours. (ii) Slaved glass transition region. Depending on the observation time and temperature, large scale motions (relaxation) are observable or appear frozen. Region ii extends from about 170K to about 210K and is centered at about 185K in 75% glycerol/water solvent. We have found a strong dependence of the glass temperature on the composition of the external solvent in our FTIR measurements. This dependence has been confirmed by the heat capacity measurements with the DSC on several different solvents with and without MbCO. Thus region ii shows a very strong dependence on external solvent. (iii) Lower equilibrium region. Transitions among the substates A₀ to A₃ occur faster than the time of observation so that the ratio r_i follows a van't Hoff relation. The temperature and pressure dependence of the ratios $r_i = A_i/A_1$ allows the energies, entropies, and volumes of the substates A_i to be determined. We find that A₀ is more tightly bound than A_1 and A_3 and that A_0 has the smallest volume and

entropy. (iv) Lower transition region. This is also an equilibrium region in which the change in slope of r_0 suggest a new transition. A possible explanation is the breaking of a few hydrogen bonds in the protein. (v) Upper equilibrium region. The region again follows a simple van't Hoff relation but the change in slope indicates that a new substate A'_0 is dominant which is less tightly bound than A_0 and which has a larger entropy and volume than substate A_1 . This second substate A'_0 may indicate an intermediate protein state to unfolding. (vi) Unfolding. The protein begins to unfold irreversibly.

Figure 2 shows the heat capacity of MbCO in 75% glycerol/water solvent as measured with a DSC. The rapid increase in the rate of heat absorption dH/dt centered at 186K is characteristic of a glass transition. No pronounced anomoly occurs until 390K (the helix-coil transition).

(b) Relaxation Experiments. In a typical pressure release experiment, a pressure of 100 MPa is applied at 240K and the sample is cooled to a temperature between 210K and 170K. The temperature is held for about 100s and the pressure is released in 10sa or less to a lower pressure (7 MPa). Infrared spectra in the CO stretching region are then taken at exponentially increasing times from 10s to 100ks.

Our main results are summarized as follows: Between 170K and 190K the width narrows and the peak frequency shifts toward the low pressure values in the case of the A_0 absorption band while the total area of A_0 remains the same, indicating internal redistribution

in the A₀ substate. This internal redistribution corresponds to transitions within tier 0 of the hierarchy of substates. Slow interconversion of A₁ and A₃ also occurs in this temperature range resulting in a concommitant change in the areas of these two bands. Fig. 3 shows the time behavior of the A bands after the fast pressure release at 180K.

Between 195K and 210K slow interconversion of A_0 with A_1 and A_3 is observed while A_1 and A_3 exchange rapidly. The transitions between different A bands correspond to transitions within tier 1 of the hierarchy. Fig. 4 shows the time behavior of the A bands after the fast pressure release at 195K. Fig. 4a gives absolute absorption spectra and Fig. 4b shows the spectra normalized to IR band A_1 . Fig. 4b indicates that A_1 and A_3 are in equilibrium at 195K.

These results show similarities to the dynamics of spin glasses and glasses: The observed relaxations are highly nonexponential in time indicating a distribution of relaxation rates. Each A substate can thus consist of many sub-substates. The relaxations "freeze out" within a narrow temperature interval of about 20K with the rates having a Vogel-Fulcher like temperature behavior in which the relaxation rates have a singularity at a temperature To significantly below the glass temperature Tg. An important difference between the glass transition in myoglobin and the glass transition in amorphous solids is the "slaving" of the transition to the composition of the external solvent in the case of proteins.

(c) Preliminary Pressure Titration Experiments. The concept of the pressure titration experiment (7,20,21) was described in the Annual Report for Year One to the Office of Naval Research. We have been conducting preliminary experiments using the microsecond, mid-infrared flash system to monitor the A₁ infrared band after photodissociation by a laser pulse. The goal of these experiments is to probe relaxation of the A₁ substate in tier 1 of the hierarchy. Freezing out of motions within tier 1 of the hierarchy of conformational substates is responsible for the nonexponential rebinding kinetics of MbCO at low temperature. In 75% glycerol/water solvent the motions in tier 1 freeze out below about 180K (13,14). Preliminary measurements are promising

3. Plans for the Third Contract Year - 1988

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Work on this contract is progressing well. We are in the process of improving the design of the pressure cell. One benefit from the improved design of the cell is a smaller thermal mass which will greatly improve the efficiency of the refrigeration system used in the pressure titration experiments. We are continuing to design and build the special equipment needed to do frequency domain studies of pressure effects. We are planning the following experiments during the third contract year.

(a) Frequency domain studies of pressure effects. The sensitivity of the CO stretching bands to pressure makes it possible to modulate the pressure with a piezoelectric crystal and to observe the corresponding pressure changes with a lock-in technique. In

principle we can extend our time (frequency) range by several orders of magnitude. We are building such a system and plan to apply it to studies of the slaved glass transition in myoglobin including observation of nonexponential relaxation at temperatures above the glass temperature $T_{\rm g}$.

- (b) Pressure titration studies of the CO stretching bands. We will continue the pressure titration studies on the A₁ band using the microsecond, mid-infrared flash photolysis system. The rather dramatic sensitivity of the intensity of the CO stretching bands makes these experiments much more feasible than in the past.
- (c) Waiting time dependence of relaxation. Relaxation studies of the magnetization in metallic spin glasses below the glass transition temperature have shown that the relaxation of the magnetization depends on the waiting time before the external field is decreased to zero. These waiting time effects have been attributed to the underlying hierarchical free energy surface (22,23) but are not fully understood. In our pressure release experiments typically the waiting times were kept at about 100s before pressure was released. We will vary the waiting time from a few seconds to about 10ks in order to determine any waiting time effects on the relaxation of the A bands.
- (d) Pressure studies on mutant myoglobins. Steve Sligar of the University of Illinois has recently succeeded in producing sperm whale myoglobin using genetic engineering (24). We have performed exploratory measurements on several mutant myoglobins

in which the distal histidine has been replaced by a different amino acid residue. We intend to extend our measurement of the infrared CO stretching bands to include the effects of pressure on the mutant myhoglobins.

(e) Ligand binding studies. Studies of the slaved glass transition and relaxation of the CO stretching bands after pressure release have proven to be unexpectedly rewarding but time-consuming. We have therefore delayed studies of the pressure dependence of ligand rebinding after flash photolysis but plan to return to them.

References

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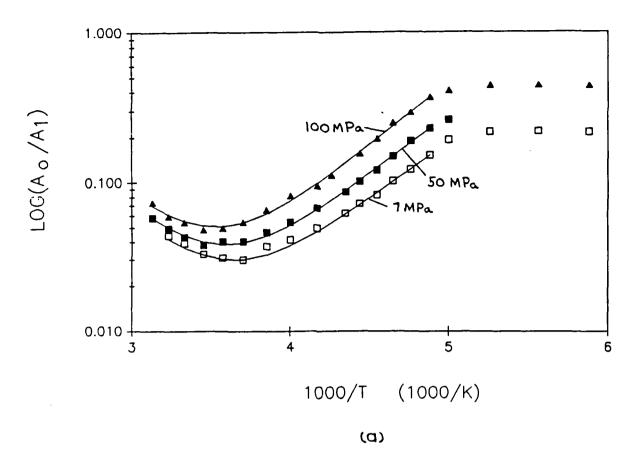
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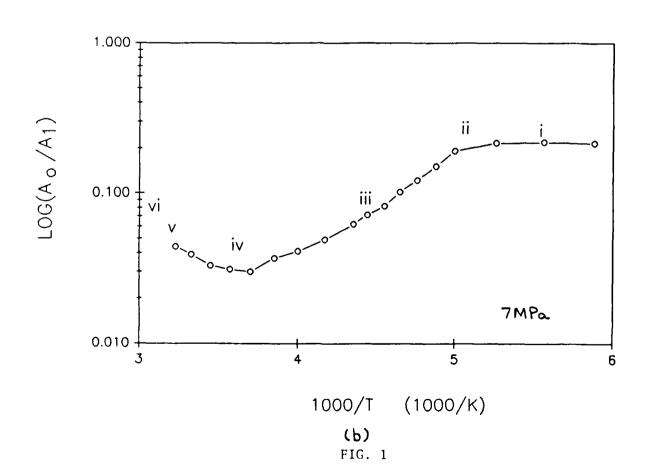
- Structure, Dynamics, and Function of Biomolecules, Ed.
 Ehrenberg, R., Rigler, R., Graslund, A., and Nilsson, L., Springer-Verlag, (1987).
- 2. Frauenfelder, H. in <u>Structure and Motion: Membranes, Nucleic Acids and Proteins</u>, Eds. Clementi, E., Corongiu, G., Sarma, M. H., and Sarma, R. H., Adenine Press, new York, p. 205, (1985).
- 3. Ansari, A. et al, Proc. Natl. Acad. Sci. USA <u>82</u>, 5000 (1985).
- 4. Beece, D. et al, Biochemistry 19, 5147 (1980).
- 5. Doster, W. et al, Biochemistry 21, 4831 (1982).
- 6. Hasinoff, B., Biochemistry, 13, 3111 (1974).
- 7. Eisenstein, L. and Frauenfelder, H., in <u>Frontiers of Biological</u>
 <u>Energetics</u>, Vol. 1, 680, Academic Press (1978).
- 8. Weber, G. and Drickamer, H., Q. Rev. Biophys. <u>16</u>, 89 (1983).
- 9. Ansari, A., Biophys. J. <u>51</u>, 289a (1987).
- *10. Ansari, A. et al, Biophys. Chem. <u>26</u>, 337 (1987).
- *11. Iben, I. E. T. et al, Biophys. J. <u>53</u>, 275a (1988).
- *12. Iben, I. E. T. et al, Biophys. J. <u>53</u>, 275a (1988).

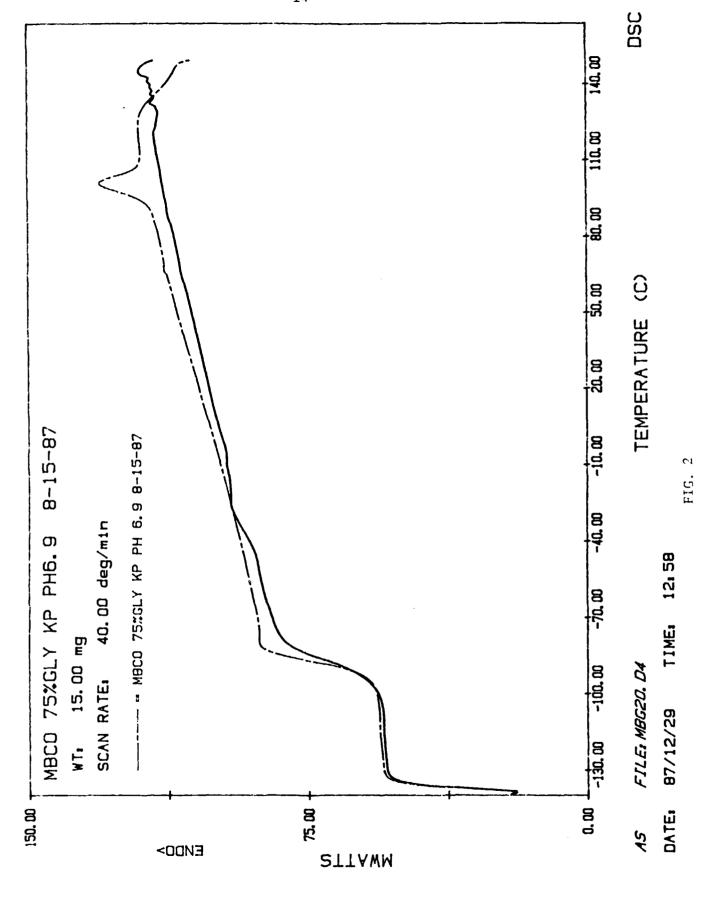
- 13. Austin, R. H. et al, Biochemistry 14, 5355 (1975).
- 14. Frauenfelder, H., Parak, F. and Young, R. D., Ann. Rev. Biophys. Biophys. Chem. 17, 451 (1988).
- 15. Stein, D., Proc. Natl. Acad. Sci. USA <u>82</u>, 0. 3670 (1985).
- 16. Binder, K. and Young, A. P., Rev. Mod. Phys. <u>58</u>, 801 (1986).
- 17. Jaeckle, J., Rep. Prog. Phys. <u>49</u>, 171 (1986).
- *18. Iben, I. E. T., Ph.D. Dissertation, University of Illinois at Urbana-Champaign (1988).
- 19. Ormos, P. et al, Biophys. J. <u>53</u>, 274a (1988).
- 20. Frauenfelder, H. and Young, R. D., Comments Mol. Cell. Biophys. 3, 347 (1986).
- 21. Young, R. D. in <u>The Time Domain in Surface and Structural Dynamics</u>, Eds. Grandjean, F. and Lang, G., Reidel Press, Dordrecht, p. 107 (1988).
- 22. Hoogerbeets, R., Luo, W.-L., Orhbach, R., Phys. Rev. <u>B34</u>, 1719 (1986).
- 23. Springer, B. and Sligar, S. G., Proc. Natl. Acad. Sci. USA <u>84</u>, 8961 (1987).

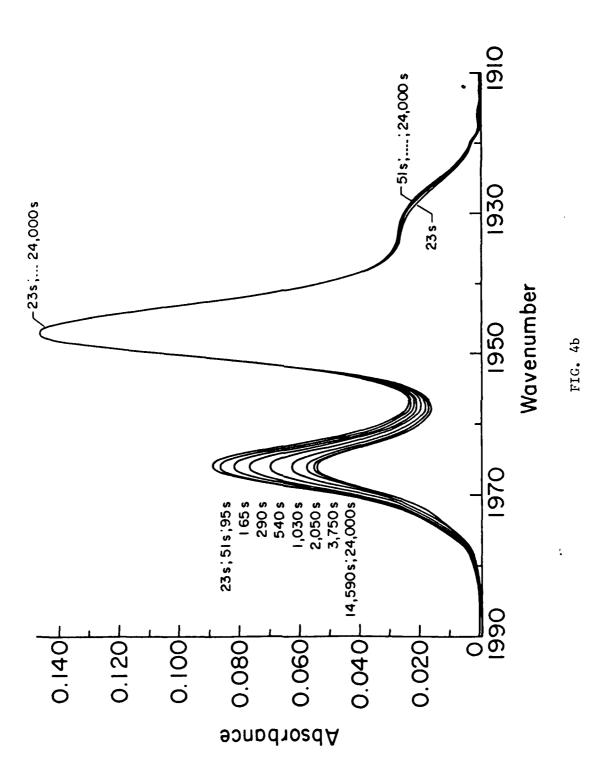
Figure Captions

- (a) Logarithm of the ratio of areas for substates A₀ and A₁ versus inverse Kelvin temperature (1000/T) for three pressures: 7 MPa, 50 MPa, and 100 MPa. Solvent: 75% glycerol/water buffered to pH 7. (b) Definitions of temperature regions at 7 MPa: (i) Glass region, (ii) Glass transition region, (iii) Lower equilibrium region, iv) Upper transition region, (v) Upper equilibrium region, (vi) Denaturation region.
- Thermal response of MbCO (75% glycerol/water buffered to pH 7 with potassium phosphate) using differential scanning calorimetry. Rate of heat absorption versus temperature (Celsius).
- Time evolution of the infrared absorption spectra corresponding to the CO stretching bands in MbCO at 180K after a fast pressure release from 100 MPa to 7 MPa. Time range: 10s to 3.8ks (75% glycerol/water buffered to pH 7 by potassium phosphate).
- Time evolution of the infrared absorption spectra corresponding to the CO stretching bands in MbCO at 195K after a fast pressure release from 100 MPa to 7 MPa. Time range: 23s to 2.4ks (75% glycerol/water buffered to pH 7 by potassium phosphate).
 - a) Absolute spectra, b) Spectra normalized to A₁.









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